



Cancer Research

Regulation of the Na⁺/H⁺ Exchanger (NHE1) in Breast Cancer Metastasis

Schammim R. Amith and Larry Fliegel

Cancer Res 2013;73:1259-1264. Published OnlineFirst February 7, 2013.

Updated version Access the most recent version of this article at:
doi:[10.1158/0008-5472.CAN-12-4031](https://doi.org/10.1158/0008-5472.CAN-12-4031)

Cited Articles This article cites by 49 articles, 24 of which you can access for free at:
<http://cancerres.aacrjournals.org/content/73/4/1259.full.html#ref-list-1>

Citing articles This article has been cited by 3 HighWire-hosted articles. Access the articles at:
<http://cancerres.aacrjournals.org/content/73/4/1259.full.html#related-urls>

E-mail alerts [Sign up to receive free email-alerts](#) related to this article or journal.

Reprints and Subscriptions To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.

Review

Regulation of the Na⁺/H⁺ Exchanger (NHE1) in Breast Cancer Metastasis

Schammim R. Amith and Larry Fliegel

Abstract

The pH gradient in normal cells is tightly controlled by the activity of various pH-regulatory membrane proteins including the isoform protein of the Na⁺/H⁺ exchanger (NHE1). NHE1 is constitutively active in a neoplastic microenvironment, dysregulating pH homeostasis and altering the survival, differentiation, and proliferation of cancer cells, thereby causing them to become tumorigenic. Cytoplasmic alkalinization in breast cancer cells occurs as a result of increased NHE1 activity and, while much is known about the pathophysiologic role of NHE1 in tumor progression with regard to ion flux, the regulation of its activity on a molecular level is only recently becoming evident. The membrane domain of NHE1 is sufficient for ion exchange. However, its activity is regulated through the phosphorylation of key amino acids in the cytosolic domain as well as by its interaction with other intracellular proteins and lipids. Here, we review the importance of these regulatory sites and what role they may play in the disrupted functionality of NHE1 in breast cancer metastasis. *Cancer Res*; 73(4); 1259–64. ©2013 AACR.

Introduction

Dysregulation of the pH gradient in and around cancer cells is a crucial step in tumor progression leading to metastasis. Abnormal increased proliferation of cells, loss of cell–cell contact, and detachment from the extracellular matrix results in a tumor metabolic microenvironment that is hypoxic, acidic, and deprived of serum (1). Extracellular acidification around tumor cells can be attributed to disrupted pH homeostasis as these cells adapt to the harsh extremes of the neoplastic milieu. Normal cells may undergo apoptosis under these conditions; however, cancer cells survive by manipulating and exploiting a host of ion exchangers including NHE1, the most common isoform of the Na⁺/H⁺ exchanger family that is ubiquitous to all mammalian cells (2). The pathophysiologic function of NHE1 in various cell types is well-documented (3), but of particular interest is that proton extrusion plays a pivotal role in cell migration, which, in cancer, is necessary for tumors to invade and metastasize at sites distant from the primary tumor (4). In breast cancer, it is suggested that dysregulation of NHE1 activity is the predominant factor leading to tumor metastasis (1); however, this claim remains to be fully substantiated. In this review, we discuss how the multiplex regulation of NHE1 activity impinges on breast cancer disease progression and current treatment regimes.

Human NHE1 is a membrane protein that is 815 amino acids in length, with residues 1 to 500 comprising the membrane

domain and residues 501 to 815 comprising the cytoplasmic tail (Fig. 1A). The membrane domain of NHE1 extrudes 1 intracellular proton in exchange for 1 extracellular sodium ion, protecting the cell from acidification and regulating intracellular pH (pH_i). Ion flux is driven by the transmembrane Na⁺ gradient and requires no direct metabolic input in normal cells. In addition, the activity of the exchanger is allosterically increased with decreasing pH_o, resulting in a rapid activation of NHE1 and a subsequent elevation of pH_i as a consequence of increased proton extrusion (5). NHE1 activity is regulated by the phosphorylation of key amino acids in its C-terminal cytosolic domain, as well as by interactions of the C-terminal tail with intracellular proteins and lipids (Fig. 1A and B; ref. 3). Regulation of NHE1 at the molecular level has been elucidated in a variety of cell types and tissues. In many cases, it is unclear if these regulatory pathways are retained in breast cancer cells, although a review of the known regulators of NHE1 can illuminate potential pathways in tumor cells. These regulators of NHE1 are activated or controlled by extracellular growth factors, hormones, and other agonists (serum, thrombin, epidermal growth factor, insulin, angiotensin II, and others; ref. 6). Ultimately, NHE1 regulators alter transport activity by altering the affinity for intracellular H⁺ such that it is more active at a more alkaline pH_i (6, 7). Much of the hormonal regulation of NHE1 is not just due to phosphorylation, but is mediated by several regulatory proteins and cofactors including calcium-binding proteins that associate with the cytosolic regulatory tail. The approximate binding sites of regulator proteins and cofactors is illustrated in Figure 1B. This includes calmodulin (CaM), which binds to a high- and low-affinity site located between amino acids 636 and 700, and blocks an autoinhibitory site, thereby activating NHE1 (8). It also includes the calcineurin homologous protein (CHP) group of regulatory calcium-binding proteins. CHP1 and CHP2 bind to NHE1 in the region of amino acids 518 to 537 (Fig. 1B), where CHP1 is

Authors' Affiliation: Department of Biochemistry, University of Alberta, Edmonton, Alberta, Canada

Corresponding Author: Larry Fliegel, University of Alberta, 347 Med Sci Bldg, Faculty Medicine, Edmonton, AB T6G 2H7, Canada. Phone: 780-492-1848; Fax: 780-492-0886; E-mail: lfliegel@ualberta.ca

doi: 10.1158/0008-5472.CAN-12-4031

©2013 American Association for Cancer Research.

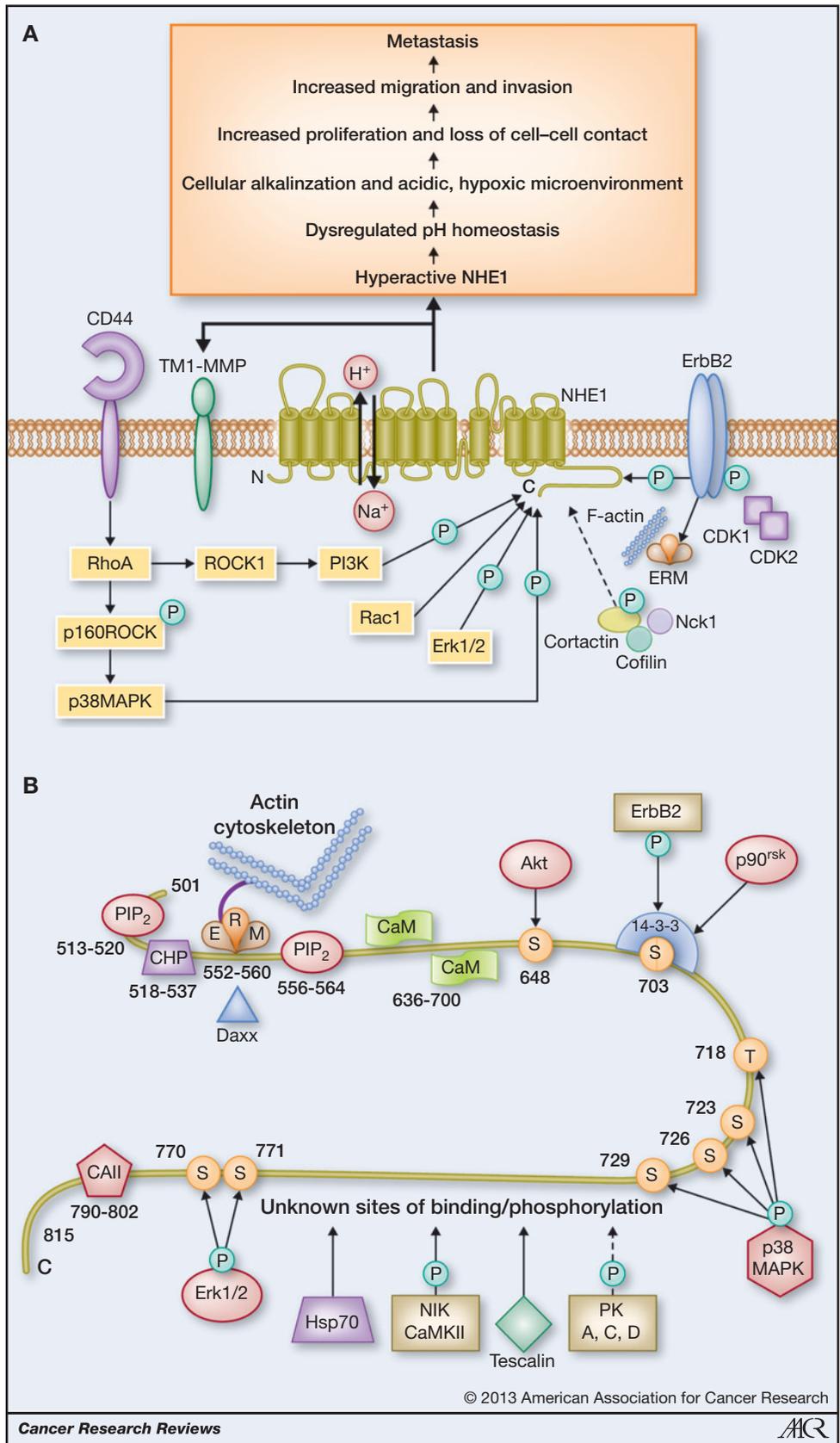


Figure 1. Schematic diagram summarizing NHE1 regulation by proteins, cofactors, and protein kinases. A, NHE1 protein in the plasma membrane with regulatory pathways in the tumor microenvironment indicated. B, the cytosolic regulatory tail of NHE1 is enlarged and indicates the location of regulatory binding sites and sites of phosphorylation. CaM, calmodulin; CHP, calcineurin homologous protein; ERM, ezrin, radixin, moesin; PIP₂, phosphatidylinositol 4,5-bisphosphate.

thought to be important for NHE1 activity and its stabilization and localization to the plasma membrane (9). CHP2 is highly expressed in tumor cells; it is protective against serum deprivation-induced cell death by increasing pH_i (10) and may play a role in the enhanced proliferation of tumor cells (11). CHP3, or tescalin (binding site uncertain), is thought to promote maturation and cell surface stability of NHE1 (12, 13). Another regulatory factor of NHE1 is phosphatidylinositol 4,5-bisphosphate (PIP₂), which binds at 2 sites between amino acids 513 and 520 and 556 and 564 (Fig. 1B). Depletion of PIP₂ results in ATP-dependent inhibition of NHE1 (14). Carbonic anhydrase II (CAII) is an enzyme that catalyzes the production of HCO₃⁻ and H⁺ from the hydration of CO₂, and it associates with NHE1 at amino acid residues 790 to 802 and increases its activity (15). In addition, the actin-binding ERM (i.e., ezrin, radixin, and moesin) proteins bind to NHE1 between amino acids 552 and 560, and direct the proper localization of NHE1 to the plasma membrane as well as maintaining cell shape (16). Daxx is a death domain-associated protein that competes with ezrin in binding to the ERM-interacting domain of NHE1 (17), while the 14-3-3 adaptor protein only binds to NHE1 when it is phosphorylated at S⁷⁰³ (Fig. 1B), thus activating NHE1 by protecting S⁷⁰³ from dephosphorylation (18). Finally, heat shock protein 70 (Hsp70) binds to NHE1 and may play a role in protein folding (19).

Several protein kinases are also involved in the regulation of NHE1 and their actions have been described in a number of studies in varying cell types. They are known to phosphorylate specific amino acids of the regulatory cytosolic domain. Figure 1B illustrates the relative position of several phosphorylation sites in this domain. Briefly, p160ROCK is a serine/threonine protein kinase and downstream target of RhoA. It facilitates the assembly of focal adhesion and actin stress fibers and was shown to mediate the RhoA activation of NHE1 in fibroblasts (20). In vascular smooth muscle cells, angiotensin II stimulates NHE1 by ERK-dependent p38MAPK phosphorylation and, in pro-B cells, NHE1-mediated apoptosis is regulated by phosphorylation at amino acids T⁷¹⁸, S⁷²³, S⁷²⁶, and S⁷²⁹ (21). Interestingly, the mutation of S^{726/729} to nonphosphorylatable alanine is protective against serum deprivation-induced cell death (22). In myocardial cells, Erk 1/2-dependent pathways are critical in the regulation of NHE1; Erk 1/2 phosphorylates NHE1 at S⁷⁷⁰ and S⁷⁷¹, while p90^{rsk}, a kinase downstream of Erk 1/2, phosphorylates NHE1 at S⁷⁰³ and, by doing so, forms a 14-3-3 ligand binding site (18). Protein kinase B (Akt) was shown to phosphorylate S⁶⁴⁸ and inhibit myocardial NHE1 activity, possibly by interfering with CaM binding (23); however, in other cell types, Akt phosphorylation stimulates NHE1 activity and is thought to be important for cell growth and survival and, possibly, in metastasis (24). Other kinases that are involved in the direct phosphorylation of NHE1, but are less defined in their site of action, include Nck-interacting kinase (NIK) and Ca²⁺/calmodulin-dependent kinase (CaMKII; refs. 25, 26). In addition, protein kinase A, C, and D are known to regulate NHE1 activity indirectly (27). Lysophosphatidic acid (LPA), a bioactive phospholipid that mediates its effects through protein kinases and by association with G-protein coupled receptors (GPCR), stimulates G_{α13}-mediated activation of both the

RhoA and Cdc42 pathways. Activation of the RhoA pathway results in p160ROCK activation, which potentiates NHE1 activation possibly by direct phosphorylation. In contrast, activation of the Cdc42 pathway mediates Erk 1/2 activation leading to NHE1 stimulation (28).

Although NHE1 plays a demonstrable role in breast cancer metastasis (outlined in Fig. 1A), the exact mechanism by which the activity of NHE1 is elevated is not well known. In breast cancer cells, early work showed that mimicking the tumor microenvironment by serum deprivation stimulates NHE1 activity in metastatic breast cancer epithelial cell lines but not in nontumorigenic cells. This was a direct result of alteration of the H⁺ affinity of the exchanger. Phosphoinositide-3-kinase (PI3K) inhibition impaired NHE1 activity in serum-supplemented conditions but potentiated the serum-deprived activation of NHE1. This upregulation of NHE1 under serum deprivation is thought to occur via a PI3K-dependent mechanism, but direct phosphorylation of NHE1 was not shown in that study (29). More recently, it was observed that in serum-deprived human breast cancer cells, the activation of NHE1 is regulated by a sequential RhoA/p160ROCK/p38MAPK signaling pathway that is gated by direct protein kinase A phosphorylation and inhibition of RhoA, suggesting that serum deprivation leads to a dynamic remodeling of the cytoskeleton forming long leading-edge pseudopodia that are uniquely poised for invasion and metastasis (30). Invadopodia, which are actin-rich invasive protrusions of these pseudopodia, are capable of proteolytic degradation of the extracellular matrix (ECM), thereby allowing invading cells to detach from the ECM and metastasize elsewhere (1). Several actin regulatory proteins including cofilin and capping protein are upregulated in these invasive cells and are specifically associated with invadopodia formation (31, 32). In MDA-MB-231 breast cancer cells, cofilin phosphorylation was found to be important in invadopodia maturation, the regulation of Nck1 binding, and cofilin activity, as well as promoting the recruitment of NHE1 to regulate pH in the invadopodia (33). CD44, a cell surface glycoprotein that serves as a receptor for hyaluronic acid (Fig. 1A), has also been shown to localize to the invadopodia of invasive breast cancer cells, stimulating NHE1 activity and invasion through the RhoA effector ROCK1. This activation of NHE1 was shown to create an acidic extracellular microenvironment that facilitates protease-mediated degradation of the extracellular matrix and promotes breast cancer cell invasion (34). The same authors also show that a RAC1 and RhoA-ROCK-PI3K pathway are involved in CD44-induced cell invasion (35), which, taken together, suggest that these signaling molecules drive NHE1 activation, cellular alkalization, and extracellular acidification, all of which facilitate invasion. Similarly, hypoxia triggers the activation of the p90 ribosomal S6 kinase (p90^{rsk}) in HT-1080 fibrosarcoma cells resulting in invadopodia formation and increased invasive capability via site-specific phosphorylation and activation of NHE1 (36), but whether or not the same is true in breast cancer cells is not known.

In a comparison between noninvasive MCF-7 cells and invasive MDA-MB-231 cells, the role of MAPK signaling pathways in NHE1-mediated breast cancer metastasis was

examined. It was found that the constitutively phosphorylated levels of Erk 1/2 and p38MAPK were higher in MDA-MB-231 cells, but both cell types expressed a similar level of phosphorylated c-Jun N-terminal kinase JNK. Treating MDA-MB-231 cells with either an NHE inhibitor (cariporide) or MAPK inhibitors suppressed the invasive capability of these cells, while treatment with a JNK-specific inhibitor had no effect (37). It is of interest to note, however, that while many studies have shown that NHE1 is regulated by MAPK in various cell types, there is also mounting evidence to suggest a role for NHE1 in the regulation of MAPK pathways (38); this could be of particular importance in the pathophysiology of breast cancer progression and could shed light on the complex interaction between the exchanger and its associated intracellular signaling molecules.

Together with the multitude of key regulatory sites on NHE1 acting as potential drug targets, several therapeutic applications may exist for the use of NHE1 inhibitors as adjuvants, to reverse dysregulated pH in the tumor environment, thereby ameliorating the efficacy of anticancer drugs on tumor growth and survival. Although there is some evidence to show that NHE1 inhibition or downregulation has an effect on the invasiveness of breast cancer cells *in vitro*, it should be noted that many studies are based on cells cultured in a well-buffered 2-dimensional (2D) milieu, generally in the absence of bicarbonate, while the tumor microenvironment *in vivo* can be different from these experimental conditions. For example, Boedtker and colleagues (39) showed that, while both Na^+/H^+ exchange and $\text{Na}^+/\text{HCO}_3^-$ cotransport contribute to pH regulation, net acid extrusion by the Na^+ , HCO_3^- cotransporter NBCn1 is the major determinant of intracellular pH regulation in primary human breast cancer carcinomas at pH_i levels less than 6.6, with NHE activity prominent under more acidic pH_i conditions (39). In spite of this, however, there is significant evidence suggesting that inhibition or deletion of NHE1 has significant effects on tumor growth in several *in vivo* models (40–43). Although these studies were not done in breast cancer cells, they are suggestive that NHE1 could play a similar role in breast cancer. Therefore, approaches to modulate NHE1 activity are of interest. Aside from targeting kinases that modulate NHE1 activity, its function could be impaired with inhibitors of various cell surface receptors, intracellular regulatory proteins, or signaling molecules, and potentially with inhibitors of extracellular agonists of the exchanger. All of these could be diverse treatment options for patients with breast cancer. One example is the receptor tyrosine kinase ErbB2 (Fig. 1A). This kinase is upregulated in approximately 30% of patients with metastatic breast cancer and is associated with resistance to paclitaxel, a chemotherapeutic agent used in the treatment of breast cancer. In a phase 2 clinical trial, ErbB2-positive mammary carcinomas became susceptible to paclitaxel treatment *in vivo* when paclitaxel was administered in combination with trastuzumab, a humanized monoclonal anti-ErbB2 antibody (44). N-terminal truncation of ErbB2 renders it constitutively active as is common in breast cancer cells where its expression is associated with increased metastatic potential. Interestingly, in MCF-7 cells expressing the truncated ErbB2 kinase, inhibition or knockdown of NHE1 sensitizes these cells to cell death

induced by the chemotherapeutic drug cisplatin. This indicates that the resultant cathepsin release may be amplified by NHE1 inhibition (45). ErbB2 upregulation is also generally associated with the inhibition of cyclin-dependent kinase CDK1 (46). In breast cancer xenograft tumors, those with significantly higher CDK1 specific activity were sensitive to paclitaxel *in vivo*, while tumors without increased CDK1 activity were resistant to the drug (47); however, there is no evidence to suggest a more direct link between CDK1 and NHE1 activity. Reshkin and colleagues, in 2003, also identified NHE1 as an essential component of paclitaxel-induced apoptosis in breast cancer cells and showed that simultaneous inhibition of NHE1 results in a concurrent enhancement of low-dose paclitaxel cell death (48). Taken together, these data indicate that the efficacy of paclitaxel and cisplatin chemotherapy may potentially be enhanced in the presence of NHE1 inhibitors.

NHE1 is also shown to mediate invasion in MDA-MB-231 cells through the regulation of membrane-type 1 matrix metalloproteinase (MT1-MMP; see Fig. 1A; ref. 37), and several MMP inhibitors exist that could potentially have a synergistic effect on decreasing invasiveness if used in combination with NHE1 inhibitors. The same may be true when using agonists or antagonists of GPCRs or inhibitors of other receptors associated with NHE1. For example, the expression of the peroxisome proliferator-activated receptor γ (PPAR γ) is greater in breast cancer cells compared with normal breast epithelium. Kumar and colleagues (49) showed that, in breast cancer cells overexpressing PPAR γ , ligand-induced activation of the receptor by its natural and synthetic agonists inhibits the proliferation of tumor cells by downregulating NHE1 transcription as well as protein expression *in vitro*. Furthermore, histopathologic analysis of breast cancer biopsies from patients treated for type 2 diabetes with the PPAR γ agonist rosiglitazone showed a marked decrease in NHE1 protein expression in the tumor tissues (49). Troglitazone, another member of the PPAR γ -ligand family, is known to induce severe acidosis mediated by the inhibition of NHE1 activity in MCF-7 and MDA-MB-231 cells, leading to a discernible reduction in cell proliferation, although this effect is highly dependent on the delivery and concentration of the drug, where it is stimulatory at low concentrations and inhibitory at higher concentrations (50). Studies like this and others highlight the need for chemotherapeutic strategies that exploit the additive anticancer effects of using drugs that inhibit NHE1 activity as well as those that target sites of NHE1 regulation.

Despite the progress made in both diagnostic and therapeutic approaches in the treatment of breast cancer, the leading cause of fatality in patients with the disease is still metastasis: the invasion and spread of the primary tumor to other sites in the body. The development of novel strategies to either inhibit tumor progression or prevent tumors from metastasizing is, therefore, essential. Because pH regulation plays such an integral role in the switch from the normal to the neoplastic microenvironment, it is imperative that the Na^+/H^+ exchanger be considered as an important target in the fight against breast cancer. With *in vitro* data suggesting an increased efficacy of chemotherapeutic drugs when used in synergy with NHE1 inhibitors, a multifaceted approach taking

into account the complex regulation of NHE1 could lead to new avenues of treatment in the search for a cure.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Authors' Contributions

Conception and design: S.R. Amith, L. Fliegel

Development of methodology: L. Fliegel

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): S.R. Amith, L. Fliegel

Writing, review, and/or revision of the manuscript: S.R. Amith, L. Fliegel
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): S.R. Amith, L. Fliegel
Study supervision: S.R. Amith, L. Fliegel

Grant Support

The research in this area (by S.R. Amith and L. Fliegel) was financially supported by the Canadian Breast Cancer Foundation.

Received October 24, 2012; revised December 14, 2012; accepted January 4, 2013; published OnlineFirst February 7, 2013.

References

- Cardone RA, Casavola V, Reshkin SJ. The role of disturbed pH dynamics and the Na⁺/H⁺ exchanger in metastasis. *Nat Rev* 2005;5:786–95.
- Neri D, Supuran CT. Interfering with pH regulation in tumours as a therapeutic strategy. *Nat Rev Drug Discov* 2011;10:767–77.
- Malo ME, Fliegel L. Physiological role and regulation of the Na⁺/H⁺ exchanger. *Can J Physiol Pharmacol* 2006;84:1081–95.
- Stock C, Cardone RA, Busco G, Krahling H, Schwab A, Reshkin SJ. Protons extruded by NHE1: digestive or glue? *Eur J Cell Biol* 2008;87:591–9.
- Wakabayashi S, Ikeda T, Iwamoto T, Pouyssegur J, Shigekawa M. Calmodulin-binding autoinhibitory domain controls "pH-sensing" in the Na⁺/H⁺ exchanger NHE1 through sequence-specific interaction. *Biochemistry* 1997;36:12854–61.
- Putney LK, Denker SP, Barber DL. The changing face of the Na⁺/H⁺ exchanger, NHE1: structure, regulation, and cellular actions. *Annu Rev Pharmacol Toxicol* 2002;42:527–52.
- Wakabayashi S, Sardet C, Fafournoux P, Counillon L, Meloche S, Pages G, et al. Structure function of the growth factor-activatable Na⁺/H⁺ exchanger (NHE1). *Rev Physiol Biochem Pharmacol* 1992;119:157–86.
- Wakabayashi S, Ikeda T, Iwamoto T, Pouyssegur J, Shigekawa M. Calmodulin-binding autoinhibitory domain controls "pH-sensing" in the Na⁺/H⁺ exchanger NHE1 through sequence specific interaction. *Biochem* 1997;36:12854–61.
- Matsushita M, Tanaka H, Mitsui K, Kanazawa H. Dual functional significance of calcineurin homologous protein 1 binding to Na(+)/H(+) exchanger isoform 1. *Am J Physiol (Cell Physiol)* 2011;301:C280–8.
- Pang T, Wakabayashi S, Shigekawa M. Expression of calcineurin B homologous protein 2 protects serum deprivation-induced cell death by serum-independent activation of Na⁺/H⁺ exchanger. *J Biol Chem* 2002;277:43771–7.
- Li QH, Wang LH, Lin YN, Chang GQ, Li HW, Jin WN, et al. Nuclear accumulation of calcineurin B homologous protein 2 (CHP2) results in enhanced proliferation of tumor cells. *Genes Cells* 2011;16:416–26.
- Li X, Liu Y, Kay CM, Muller-Esterl W, Fliegel L. The Na(+)/H(+) exchanger cytoplasmic tail: Structure, function, and interactions with tescalcin. *Biochemistry* 2003;42:7448–56.
- Zaun HC, Shrier A, Orlowski J. Calcineurin B homologous protein 3 promotes the biosynthetic maturation, cell surface stability, and optimal transport of the Na⁺/H⁺ exchanger NHE1 isoform. *J Biol Chem* 2008;283:12456–67.
- Aharonovitz O, Zaun HC, Balla T, York JD, Orlowski J, Grinstein S. Intracellular pH regulation by Na(+)/H(+) exchange requires phosphatidylinositol 4,5-bisphosphate. *J Cell Biol* 2000;150:213–24.
- Li X, Liu Y, Alvarez BV, Casey JR, Fliegel L. A novel carbonic anhydrase II binding site regulates NHE1 activity. *Biochemistry* 2006;45:2414–24.
- Denker SP, Huang DC, Orlowski J, Furthmayr H, Barber DL. Direct binding of the Na–H exchanger NHE1 to ERM proteins regulates the cortical cytoskeleton and cell shape independently of H(+) translocation. *Mol Cell* 2000;6:1425–36.
- Jung YS, Kim HY, Kim J, Lee MG, Pouyssegur J, Kim E. Physical interactions and functional coupling between Daxx and sodium hydrogen exchanger 1 in ischemic cell death. *J Biol Chem* 2008;283:1018–25.
- Lehoux S, Abe J, Florian JA, Berk BC. 14–3–3 Binding to Na⁺/H⁺ exchanger isoform-1 is associated with serum-dependent activation of Na⁺/H⁺ exchange. *J Biol Chem* 2001;276:15794–800.
- Silva NL, Haworth RS, Singh D, Fliegel L. The carboxyl-terminal region of the Na⁺/H⁺ exchanger interacts with mammalian heat shock protein. *Biochemistry* 1995;34:10412–20.
- Tominaga T, Ishizaki T, Narumiya S, Barber DL. p160ROCK mediates RhoA activation of Na–H exchange. *EMBO J* 1998;17:4712–22.
- Khaled AR, Moor AN, Li A, Kim K, Ferris DK, Muegge K, et al. Trophic factor withdrawal: p38 mitogen-activated protein kinase activates NHE1, which induces intracellular alkalinization. *Mol Cell Biol* 2001;21:7545–57.
- Grenier AL, Abu-ihweij K, Zhang G, Ruppert SM, Boohaker R, Slepko ER, et al. Apoptosis-induced alkalinization by the Na⁺/H⁺ exchanger isoform 1 is mediated through phosphorylation of amino acids Ser726 and Ser729. *Am J Physiol Cell Physiol* 2008;295:C883–96.
- Snabaitis AK, Cuello F, Avkiran M. Protein kinase B/Akt phosphorylates and inhibits the cardiac Na⁺/H⁺ exchanger NHE1. *Circ Res* 2008;103:881–90.
- Meima ME, Webb BA, Witkowska HE, Barber DL. The sodium–hydrogen exchanger NHE1 is an Akt substrate necessary for actin filament reorganization by growth factors. *J Biol Chem* 2009;284:26666–75.
- Yan W, Nehrke K, Choi J, Barber DL. The Nck-interacting kinase (NIK) phosphorylates the Na⁺-H⁺ exchanger NHE1 and regulates NHE1 activation by platelet-derived growth factor. *J Biol Chem* 2001;276:31349–56.
- Fliegel L, Walsh MP, Singh D, Wong C, Barr A. Phosphorylation of the carboxyl-terminal domain of the Na⁺/H⁺ exchanger by Ca²⁺/calmodulin-dependent protein kinase II. *Biochem J* 1992;282:139–45.
- Loo SY, Chang MK, Chua CS, Kumar AP, Pervaiz S, Clement MV. NHE-1: a promising target for novel anti-cancer therapeutics. *Curr Pharm Des* 2012;18:1372–82.
- Hooley R, Yu CY, Symons M, Barber DL. G alpha 13 stimulates Na⁺-H⁺ exchange through distinct Cdc42-dependent and RhoA-dependent pathways. *J Biol Chem* 1996;271:6152–8.
- Reshkin SJ, Bellizzi A, Albarani V, Guerra L, Tommasino M, Paradiso A, et al. Phosphoinositide 3-kinase is involved in the tumor-specific activation of human breast cancer cell Na(+)/H(+) exchange, motility, and invasion induced by serum deprivation. *J Biol Chem* 2000;275:5361–9.
- Cardone RA, Bagorda A, Bellizzi A, Busco G, Guerra L, Paradiso A, et al. Protein kinase A gating of a pseudopodial-located RhoA/ROCK/p38/NHE1 signal module regulates invasion in breast cancer cell lines. *Mol Biol Cell* 2005;16:3117–27.
- Wang W, Eddy R, Condeelis J. The cofilin pathway in breast cancer invasion and metastasis. *Nat Rev Cancer* 2007;7:429–40.
- Ayala I, Baldassarre M, Giacchetti G, Caldieri G, Tete S, Luini A, et al. Multiple regulatory inputs converge on cortactin to control invadopodia biogenesis and extracellular matrix degradation. *J Cell Sci* 2008;121(Pt 3):369–78.
- Magalhaes MA, Larson DR, Mader CC, Bravo-Cordero JJ, Gil-Henn H, Oser M, et al. Cortactin phosphorylation regulates cell invasion through a pH-dependent pathway. *J Cell Biol* 2011;195:903–20.
- Bourguignon LY, Singleton PA, Diedrich F, Stern R, Gilad E. CD44 interaction with Na⁺-H⁺ exchanger (NHE1) creates acidic

- microenvironments leading to hyaluronidase-2 and cathepsin B activation and breast tumor cell invasion. *J Biol Chem* 2004;279:26991-7007.
35. Bourguignon LY, Zhu H, Shao L, Chen YW. CD44 interaction with tiam1 promotes Rac1 signaling and hyaluronic acid-mediated breast tumor cell migration. *J Biol Chem* 2000;275:1829-38.
 36. Lucien F, Brochu-Gaudreau K, Arsenault D, Harper K, Dubois CM. Hypoxia-induced invadopodia formation involves activation of NHE-1 by the p90 ribosomal S6 kinase (p90RSK). *PLoS One* 2011;6:e28851.
 37. Lin Y, Chang G, Wang J, Jin W, Wang L, Li H, et al. NHE1 mediates MDA-MB-231 cells invasion through the regulation of MT1-MMP. *Exp Cell Res* 2011;317:2031-40.
 38. Pedersen SF, Darborg BV, Rentsch ML, Rasmussen M. Regulation of mitogen-activated protein kinase pathways by the plasma membrane Na⁺/H⁺ exchanger, NHE1. *Arch Biochem Biophys* 2007;462:195-201.
 39. Boedtker E, Moreira JM, Mele M, Vahl P, Wielenga VT, Christiansen PM, et al. Contribution of Na⁺,HCO₃⁻-cotransport to cellular pH control in human breast cancer: a role for the breast cancer susceptibility locus NBCn1 (SLC4A7). *Int J Cancer J* 2012. In Press.
 40. Lagarde AE, Franchi AJ, Paris S, Pouyssegur JM. Effect of mutations affecting Na⁺:H⁺ antiport activity on tumorigenic potential of hamster lung fibroblasts. *J Cell Biochem* 1988;36:249-60.
 41. Rotin D, Steele-Norwood D, Grinstein S, Tannock I. Requirement of the Na⁺/H⁺ exchanger for tumor growth. *Cancer Res* 1989;49:205-11.
 42. Reshkin SJ, Bellizzi A, Caldeira S, Albarani V, Malanchi I, Poignee M, et al. Na⁺/H⁺ exchanger-dependent intracellular alkalinization is an early event in malignant transformation and plays an essential role in the development of subsequent transformation-associated phenotypes. *FASEB J* 2000;14:2185-97.
 43. Yamagata M, Tannock IF. The chronic administration of drugs that inhibit the regulation of intracellular pH: *in vitro* and anti-tumour effects. *Br J Cancer* 1996;73:1328-34.
 44. Slamon DJ, Leyland-Jones B, Shak S, Fuchs H, Paton V, Bajamonde A, et al. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *New Engl J Med* 2001;344:783-92.
 45. Lauritzen G, Stock CM, Lemaire J, Lund SF, Jensen MF, Damsgaard B, et al. The Na⁺/H⁺ exchanger NHE1, but not the Na⁺, HCO₃⁻ cotransporter NBCn1, regulates motility of MCF7 breast cancer cells expressing constitutively active ErbB2. *Cancer Lett* 2012;317:172-83.
 46. Castedo M, Perfettini JL, Roumier T, Kroemer G. Cyclin-dependent kinase-1: linking apoptosis to cell cycle and mitotic catastrophe. *Cell Death and Differentiation* 2002;9:1287-93.
 47. Nakayama S, Torikoshi Y, Takahashi T, Yoshida T, Sudo T, Matsushima T, et al. Prediction of paclitaxel sensitivity by CDK1 and CDK2 activity in human breast cancer cells. *Breast Cancer Res* 2009;11:R12.
 48. Reshkin SJ, Bellizzi A, Cardone RA, Tommasino M, Casavola V, Paradiso A. Paclitaxel induces apoptosis via protein kinase A- and p38 mitogen-activated protein-dependent inhibition of the Na⁺/H⁺ exchanger (NHE) NHE isoform 1 in human breast cancer cells. *Clin Cancer Res* 2003;9:2366-73.
 49. Kumar AP, Quake AL, Chang MK, Zhou T, Lim KS, Singh R, et al. Repression of NHE1 expression by PPARgamma activation is a potential new approach for specific inhibition of the growth of tumor cells *in vitro* and *in vivo*. *Cancer Res* 2009;69:8636-44.
 50. Turturro F, Friday E, Fowler R, Surie D, Welbourne T. Troglitazone acts on cellular pH and DNA synthesis through a peroxisome proliferator-activated receptor gamma-independent mechanism in breast cancer-derived cell lines. *Clin Can Res* 2004;10:7022-30.